Targeted Protein Degradation Drug Discovery Solutions





Powering the Future of Degrader Therapeutics

Targeted Protein Degradation (TPD) is an innovative strategy that removes disease-related proteins directly, overcoming limitations of traditional inhibitors that merely block protein function. By harnessing the cell's own degradation machinery, TPD offers new therapeutic solutions for previously "undruggable" targets.

At ICE Bioscience, we specialize in supporting the development of these leading TPD modalities:

- PROTACs
- Molecular Glues
- DACs (Degrader-Antibody Conjugates)



Figure 1. (A) A bifunctional molecule that links a target protein to an E3 ligase, inducing ubiquitination and degradation. (B) A small molecule that enhances the interaction between an E3 ligase and a target protein, promoting degradation. (C) DACs are made of an antibody for precise targeting, chemically linked to a degrader that induces protein degradation in specific cells.

Why Partner with Us?



 Seamless workflow: From in-house protein expression to complex assays, ensuring seamless integration.



• Comprehensive degradation analysis: High-throughput HiBiT assays and validation with Jess, ICW, and other classic methods.



• Mass spectrometry-based proteomics for TurboID labeling and protein degradation profiling.

1000+ Global Partnerships



• Diverse phenotypic screening tools for functional evaluation.

IMPACT IN NUMBERS



400+ Employees



15 Years of Excellence



10000+ Studies Annually



Integrated TPD Screening Workflow



¹ In some cases, a degradation-first screening strategy may be applied to quickly assess target degradation before detailed binding and complex formation studies.

Your TPD Starts Here: 700+ Ready-to-Use Proteins

- 700+ purified proteins, including 350+ kinases
- Specific activity and lot-to-lot consistency
- Covering Top TPD targets
 - \checkmark KRAS and mutants
 - √ VAV1
 - √ NEK7
 - √ CDK2
 - $\checkmark~$ BTK and mutants
 - √ c-Myc
 - $\checkmark~$ EGFR and mutants
 - √ IRAK4
 - √ HPK1
 - \checkmark BRD4
 - √ STAT6
 - √ GSPT1
 - √ IKZF1
 - √ PLK3
 - ...



Target Validation

Target validation is essential in TPD drug discovery to assess whether a protein is biologically relevant and susceptible to degradation. We provide comprehensive validation services to confirm target feasibility, degradability, and therapeutic potential, supporting efficient TPD research.

Assay Category	Assay Type	Description and Purpose	
Genetic Modulation	- CRISPR/Cas9 Knockout (KO)	Uses CRISPR or RNAi to reduce or eliminate target protein expression, confirming its essentiality for cellular functions or disease pathways.	
	- RNA Interference (RNAi)		
	- qPCR	Evaluates the mRNA and protein levels of the target and determines its subcellular or tissue localization, validating its relevance in specific biological contexts.	
Expression and Localization	- Western Blotting		
	- Immunohistochemistry (IHC)		
Protein Production	- Recombinant Protein Expression and Purification	Supplies high-quality proteins for use in binding assays, complex formation studies, or functional and mechanistic research.	
Protein Stability and Turnover	- Protein t1/2 Assay	Measures the natural degradation rate (half-life) or ligand-induced stability changes of the target protein to assess its suitability for therapeutic degradation strategies.	
	- Thermal Shift Assay (TSA)/CETSA		
Functional Dependency	- Cell Viability Assays	Evaluates how modulating the target protein affects survival, proliferation,	
	- Reporter Gene Assays	phenotypes.	

TurbolD Proximity Proteomics

TurboID proximity labeling combined with quantitative proteomics provides a powerful platform for mapping protein interaction networks and uncovering the mechanisms of TPD. By biotinylating proteins in close proximity to a bait protein in living cells, TurboID captures dynamic protein interactions under physiological conditions, allowing downstream mass spectrometry to identify and quantify associated partners.



DIA Proteomics for Substrate Discovery

Our DIA (Data-Independent Acquisition)-based proteomics service combines automated, high-throughput sample prep with the ultra-fast, high-resolution Orbitrap Exploris™ 480 or Astral MS platform, enabling early identification of degrader substrates. From large-scale screening to precise degradation profiling, we deliver deep, reliable proteome insights—fast.



Multi-depth Data Analysis

Wet-lab Validation

Binary and Ternary Complex Formation Assays

Complex formation assays enable comprehensive characterization of binary interactions between proteins and ligands, as well as ternary complex formation with E3 ligases.

• Biochemical Assays: Using techniques such as TR-FRET, we evaluate binary interactions between protein-of-interest (POI) and ligands or binders.



Figure 2. Complexes are formed among target-PROTAC-E3 ligase, with Tb as the donor fluorophore and AF488 as the acceptor fluorophore. A typical bell-shaped dose-response curve is shown.



Figure 3. TR-FRET dose-response curves for binary and ternary complex formation in a molecular glue degrader (MGD) system.

• Biophysical Assays: Advanced tools like Spectral Shift (SpS), Temperature-Related Intensity Change (TRIC) and SPR, allow us to characterize the kinetic and thermodynamic properties of binding events, offering mechanistic insights into complex formation.



Figure 4. Spectral Shift assays demonstrate the binding affinity of a PROTAC to CRBN (binary complex, left) and the formation of a ternary complex with STAT6 (right).

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· Cell-based Assays: Cell-based NanoBRET and NanoBiT assays enable the validation of binary and ternary interactions in their native context.



Figure 5. NanoBRET measures ternary complex formation by detecting energy transfer between luminescent and fluorescent tags on proteins, indicating their proximity. This assay shows the concentration-dependent formation of the IRAK4 ternary complex in the presence of MG-132.

Protein Degradation Assays

HiBiT-Based In Vitro Lytic Assays

HiBiT, an 11-amino acid tag from NanoLuc® luciferase, enables real-time detection of PROTAC and Molecular Glue -induced protein degradation. Upon target degradation, HiBiT signal decreases as it no longer binds LgBiT to form an active luciferase. Integrated via CRISPR/Cas9 or transient expression, HiBiT provides a quantitative, high-throughput solution for evaluating degrader efficacy in live cells.



Figure 6. HiBiT-based endogenous tagging and luminescent detection of target proteins.





DIA Proteomics for Off-target Profiling

Our DIA-based proteomics enables quantitative and unbiased off-target analysis for molecular glue degraders and PROTACs, facilitating early identification of unintended substrate interactions.



Classic Protein Degradation Assays

Protein degradation assays deliver quantitative, reproducible data to confirm target degradation, evaluate dose/time responses, and detect off-target effects. By integrating high-sensitivity, high-throughput technologies, our assays accelerate target validation, degrader optimization, and drug development.

Category	Assays	
Traditional Protein Detection	WB, Jess (Simple Western), ICW	
Homogeneous HTS	HTRF, AlphaLISA	
Multiplex & High-Sensitivity	MSD, Flow Cytometry	
Imaging-Based Assays	HCA (High Content Analysis)	



Figure 7: Time-dependent degradation of VAV1 in human PBMCs measured by Jess.

Confirm the Mechanism Behind Your Degrader

Understanding how your degrader works is critical for optimizing efficacy, selectivity, and therapeutic impact. Our comprehensive MOA services provide key insights into ubiquitination, degradation pathways, and functional consequences.

Question	Our Approach	
Is the POI ubiquitinated?	WB, PRM-MS, in vitro ubiquitination assay	
Does degradation rely on the proteasome?	Proteasome Inhibitor Assays (MG-132), CHX Chase	
What are the kinetics of degradation?	HiBiT Assay, MSD, ICW, Time-Course WB	
What happens after POI degradation?	Phosphoproteomics, Pathway Assays, Phenotypic Analysis	

From Target Engagement to Phenotypic Impact



DMPK and In Vivo Pharmacology

Our DMPK service assays provide comprehensive support for PROTAC discovery, lead optimization, clinical candidate selection, and IND filing processes.

	Lead Discovery and Optimization	Clinical Candidate Selection	Clinical Candidate Characterization and IND Filing
Biotransformation	 Metabolic Soft Spot Analysis Reactive Metabolite Screening Special Metabolizing Enzyme Phenotyping 	 Metabolism in Hepatocytes Across Species ADME in BDC Rats Radiolabeled Tissue Distribution Study in Rats 	 Radiolabeled ADME Study in Animals Radiolabeled Tissue Destruction Study Metabolizing Enzyme Mapping
In Vitro DDI/ADME	 Solubility and Permeability P-gp/BCRP Substrate Analysis Plasma Protein Binding Metabolic Stability 	CYP Inhibition and Induction Transporter Inhibition	Transporter Substrate Analysis
Bioanalysis/PK	PK Screening in Animals PK/PD in Pharmacological Model	 PK/TK in Animals Biomarker Exploration Tissue Distribution Studies of the Parent Excretion Studies of the Parent 	 Full PK Studies in Animals Tissue Distribution Studies of the Parent Excretion Studies of the Parent

 Cancer Models Neurodegenerative Disease Models Inflammatory and Autoimmune Disease Models
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 Inflammatory and Autoimmune Disease Models
Disease Model Development and Customization

In Vivo Mechanistic Studies
In Vivo Target Degradation
Pathway Analysis
Tissue Distribution Studies

Biomarker Development

Decoding the Mechanism and Impact of a VAV1 Molecular Glue

Background

VAV1 is a key signaling protein in T cell activation and immune regulation, with aberrant expression linked to autoimmune diseases and cancer. Despite its biological relevance, VAV1 has long been considered undruggable. Recent advances in molecular glue degraders (MGDs) have enabled selective targeting of such proteins. Building on this concept, our study aims to evaluate a VAV1-directed MGD using a suite of *in vitro* and *in vivo* assays to characterize its degradation efficiency, mechanism of action, and functional impact.

Results

This case study uses a non-disclosed VAV1 degrader compound to showcase the capabilities of our assay platforms for molecular glue evaluation.

1. VAV1 Binary and Ternary Complex Formation



Figure 8: Evaluation of VAV1-targeting molecular glue complex formation. Left: Ternary complex formation across VAV1 proteins from different species, detected via HTRF. Middle: Spectral shift assay showing ternary binding with a determined Kd of 16.3 nM. Right: Summary of binary and ternary complex formation using recombinant human VAV1, assessed by HTRF, spectral shift, and NanoBRET PPI assays.

2. VAV1 Degradation Assays

2.1 In vitro HiBiT Assays and Simple Western Assays in Primary Immune Cells



Figure 9: Quantification of VAV1 degradation across cell types. Left: Representative protein detection by JESS showing dose-dependent VAV1 degradation across multiple cell types. Middle: Dose-response curves in Jurkat cells show strong agreement between HiBiT (red) and JESS (blue) platforms. Right: Summary of DC_{50} values derived from HiBiT and JESS assays, indicating potent and consistent degradation across multiple immune cell types.





Figure 10: *In vivo* degradation of mouse VAV1 (A) Flow cytometry detection of VAV1 levels in CD3⁺ T cells from mouse whole blood and splenocytes 24 hours after oral dosing with VAV1 MGD. MFI reduction indicates effective *in vivo* degradation. (B) Jess-based quantification of VAV1 in mouse white blood cells and splenocytes confirms protein level reduction at 10 mg/kg dosing.

3. Functional Assays in Primary Immune Cells

To evaluate the immunomodulatory effects of the VAV1 degrader, we conducted functional assays in primary immune cells. These assays provide insights into the compound's impact on immune cell function and inflammatory signaling.

Complex Type	Functional Assay	IC ₅₀ (nM)
T cells	Proliferation (Flow cytometry)	0.35
	CD69 Detection (Flow cytometry)	0.32
	IL-2 Secretion	0.35
B cells	CD69 Detection (Flow cytometry)	0.77
	IL-6 Secretion	0.75
Th17 polarization	IL-17A Secretion (ELISA)	1.95

4. MOA Studies

We provide rapid, multi-angle analysis of degradation mechanisms using CHX chase, time-course assays, and pathway validation to confirm proteasome dependency and degrader selectivity.



Figure 11: VAV1 undergoes dose- and time-dependent degradation. Left: CHX chase assay confirms active protein turnover. Right: Time- and dose-dependent degradation kinetics of VAV1 measured by HiBiT assay.



Figure 12: VAV1 degradation is mediated by the proteasome pathway. Treatment with the VAV1 molecular glue induces significant degradation of VAV1 protein, which is rescued by the proteasome inhibitor MG132. This indicates that VAV1 is degraded via a proteasome-dependent mechanism.

5. DIA-MS Off-Target Profiling

Unbiased, high-depth proteomic profiling to identify on- and off-target effects of degraders. Quantify thousands of proteins per sample across time points and doses to support selectivity assessment and mechanism validation.



Figure 13. DIA-based proteomic profiling and Western blot confirm selective VAV1 degradation in human PBMCs. PBMCs were treated with VAV1 MGD (50 nM and 10 µM, 24 h). DIA profiling showed significant VAV1 degradation, with minor effects on VAV2 and VAV3. Western blot (right) validated these results.

VCE Innovative GRO⁺Explorer ICE Bioscience

ICE Bioscience was founded in 2010 as an Innovative CRO+ Explorer company. We specialize in early drug discovery services, spanning from target validation to the identification of pre-clinical candidates. We stand out for our collaborative spirit and expertise in boldly exploring new therapeutic target research. Our commitment to drug discovery services, delivered with enthusiasm and professionalism, empowers clients to overcome challenges, address scientific puzzles, and fulfill our promises to clients, communities, the environment, and global health.



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